

# Is Pepsin a Reliable Marker of Laryngopharyngeal Reflux? A Systematic Review

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## Abstract

**Objective.** Laryngopharyngeal reflux (LPR) is a common illness of otolaryngology visits. Over the past few years, pepsin has become a promising marker of LPR. The objective of the present research is to analyze the existing literature using pepsin as a diagnostic tool of LPR through a systematic review.

**Data Sources.** PubMed (Medline), Trip Database, Cochrane Library, EMBASE, SUMsearch, and Web of Science.

**Review Methods.** The outcome assessed was the presence of pepsin in LPR patients. We included articles in which pepsin was studied in LPR patients (clinically suspected or with confirmed diagnosis). Studies with no control group, comparison group, and/or a sample size lower than 20 patients were excluded.

**Results.** Twelve studies were included. All included studies, with the exception of 2, found statistically significant differences for pepsin in cases compared with healthy controls.

**Conclusion.** Pepsin might be a reliable marker in LPR patients, although questions remain about optimal timing, location, nature, and threshold values for pepsin testing. Future investigations are necessary to clarify the best method to use pepsin in the diagnostic process of LPR.

## Keywords

extraesophageal reflux, laryngopharyngeal reflux, LPR, pepsin, systematic review, reflux

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Patients with gastroesophageal reflux (GERD) often display classic symptoms such as pyrosis or regurgitation, but other atypical symptoms, such as chronic cough, asthma, or laryngitis, may appear. These atypical symptoms can be identified as laryngopharyngeal reflux (LPR).<sup>1</sup>

LPR accounts for 4% to 10% of patients seen in otolaryngology consultations. It usually has nonspecific symptoms, making its diagnosis difficult. An important issue contributing to this difficult diagnosis is the lack of reliable diagnostic tests. Multichannel intraluminal impedance (MII) and 24-hour dual-probe pH-metry are considered the gold standard.<sup>2</sup> The reproducibility of pH-metry is poor<sup>3</sup> as its outcome is dependent on the position of the proximal sensor, and the pH cutoff value is not clear yet.<sup>4</sup> In addition, these tests cannot be performed in all patients suspected of presenting LPR given their invasive nature and high cost.

Instead of these diagnostic tools, otolaryngologists usually rely on clinical questionnaires such as the Reflux Symptom Index (RSI),<sup>5</sup> tests such as Reflux Finding Score (RFS),<sup>6</sup> and a proton-pump inhibitor (PPI) therapeutic trial<sup>7</sup> to make a diagnosis. There are some limitations for these diagnostic tests.<sup>8–10</sup>

The result of these limitations is that LPR is usually misdiagnosed. For this reason, the economic burden of misdiagnosing LPR has been highlighted by many authors.<sup>11–14</sup>

The gastric enzyme, pepsin, has been proposed as a solution to overcome the aforementioned limitations. Pepsin is a proteolytic enzyme that is secreted by the stomach's chief cells as the zymogen pepsinogen. As there is no production of pepsin in the airway, its presence is a sign of LPR.

Some investigations have demonstrated the biologic plausibility of pepsin taking part in LPR,<sup>1,15–21</sup> and in recent

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**Table 1.** Quality Score.

| Item Assessed  | Characteristic                                    | Weight |
|--|---|--------|
| Sample size  | >70   | 2      |
|  | 40-69   | 1      |
|  | 20-39   | 0      |
| Gold-standard comparison                             | Multichannel intraluminal impedance or dual probe | 2      |
|  | pH monitoring                                     | 1      |
|  | None or others                                    | 0      |
| Blinded study  | Yes   | 2      |
|  | No  | 0      |
| Time between gold-standard test and pepsin detection | <1 week   | 2      |
|  | >1 week   | 0      |
| Significance of the sample                           | Yes   | 2      |
|  | Partially   | 1      |
|  | No  | 0      |
| Total  |   |        |

years, an increasing number of investigations have attempted to demonstrate that pepsin is a reliable marker for LPR. Nevertheless, different methods have been used to compare and analyze the available evidence, and results have been contradictory.

The objective of the present research article is to analyze the existing literature using pepsin as a diagnostic tool of LPR through a systematic review.

## Methods

### Literature Search: Inclusion and Exclusion Criteria

Recommendations of the PRISMA statement were followed. We performed a systematic review using PubMed (Medline), Trip Database, Cochrane Library, EMBASE, SUMsearch, and Web of Science. We used a predefined search strategy employing a combination of keywords (*pepsin, laryngopharyngeal reflux, saliva, sputum, nasal, airway, GERD, extraesophageal reflux*) complemented with free text terms. We also manually reviewed the reference lists of all selected articles to identify studies potentially fulfilling inclusion criteria but not found by the initial search method.

The outcome assessed was the presence of pepsin in LPR patients. The title and abstracts of the studies retrieved were thoroughly reviewed, and those fulfilling inclusion criteria were independently assessed for eligibility by 2 review team members (C.C.-H. and P.V.). Any disagreement between them was resolved through discussion with a third reviewer (A.R.-R.).

We have included studies written in both English and Spanish. There were no restrictions by date or publication type, and the last update of the search was performed in December 2016.

We included articles in which pepsin was studied in LPR patients (clinically suspected or with confirmed diagnosis).

Articles were included even if pepsin measurement was not the main objective of the work. Studies with no comparison group and/or a sample size lower than 20 patients were excluded.

### Assessment of the Study Quality

Due to the heterogeneity of the selected works, we developed a score to assess the quality and the risk of bias of the included studies.

The scale had 5 items, with a score range from 0 to 10 points. The items considered were the following: sample size, gold-standard comparison, blinded study, time lapse between gold-standard test and pepsin detection, and significance of the sample. Articles were included regardless of its score. The scale with the scoring of each item appears in **Table 1**.

## Results

### Search Results

A description of the research process appears in **Figure 1**. We obtained 146 studies, and after reading all the abstracts, 28 were selected for full-text reading. Of them, 12 fulfilled the established inclusion criteria.

Of the selected articles for full-text reading, 5 were excluded because no control group was present, 4 did not study LPR patients, 2 did not study the association between LPR and pepsin, 2 were written in Chinese, and 3 had a sample size lower than 20 patients.

### Results of the Included Studies

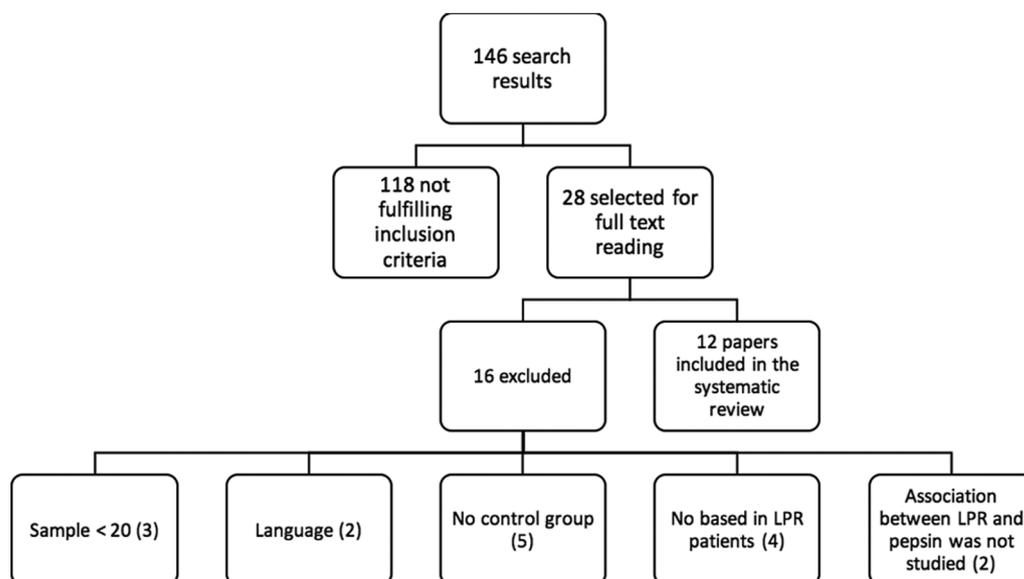
The description of included results is shown in **Table 2**.

All included studies, with the exception of 2,<sup>22,23</sup> found statistically significant differences for pepsin in diagnosed cases compared with healthy controls.

Yadlapati et al<sup>22</sup> studied a sample of 33 participants distributed in 3 cohorts: asymptomatic controls, laryngeal symptoms, and laryngeal and esophageal symptoms. They tested for pepsin in saliva using the Peptest (RD Biomed, East Yorkshire, England) did not find statistically significant differences. However, when an estimation of pepsin concentration in each cohort was performed, statistically significant differences were then found. Differences in favor of laryngeal and esophageal cohorts symptoms were  $P = .01$  and  $.04$ .

Komatsu et al<sup>23</sup> studied a sample of 55 patients with LPR symptoms. For the control group, 10 patients with typical GERD symptoms (no LPR symptoms) were randomly selected. This study did not find differences in pepsin concentration, but unlike the other studies included in this review, they used GERD patients, asymptomatic for LPR, and not healthy patients as the control group.

The most prominent study published to date regarding quality scores was performed by Na et al,<sup>24</sup> with a score of 9 points. This work is also the study with the second largest sample size of LPR patients, including 62 participants, 50 confirmed LPR patients, and 12 asymptomatic controls. They looked for pepsin in saliva via enzyme-linked immunosorbent assay (ELISA). Samples were obtained at



**Figure 1.** Flow diagram depicting the identification and selection of eligible studies for inclusion in the systematic review. LPR, laryngopharyngeal reflux.

different times of the day. They found higher levels of pepsin in patients than in controls. They also found higher levels of pepsin upon waking, whereupon they highlighted the importance of taking samples upon waking.

The study with the largest sample size was performed by Fortunato et al,<sup>25</sup> with 133 pediatric patients. This work and the one performed by Iannella et al<sup>26</sup> were the only studies performed in pediatric patients. Both used patients with a confirmed diagnosis of LPR.

There is controversy among the selected studies about the presence of pepsin in healthy controls. Eight of the included studies found pepsin in healthy controls,<sup>2,22,24,25,27-30</sup> while another 3 did not.<sup>20,26,31</sup> In the group of studies that did not find pepsin in healthy controls, 2 of 3 used controls confirmed via gold-standard probes.<sup>20,31</sup> On the other hand, the whole group of studies that found pepsin in controls used clinically healthy controls, not confirmed controls.

With regard to the samples to be examined, studies can be divided into 2 main groups: 9 studied fluids,<sup>2,22,24-28,30,31</sup> whereas the other 3 studied biopsies.<sup>20,23,29</sup>

Within the studies that sampled biopsies, the work of Jiang et al<sup>29</sup> stands out as the only one to study pepsin in biopsies taken from awakened patients. The study of pepsin using immunohistochemistry (IHQ) was first described by Johnston et al,<sup>20</sup> but only in anesthetized patients.

Jian et al<sup>29</sup> studied a sample of 15 confirmed LPR patients and 21 controls. Biopsies were taken from interarytenoid mucosa. The difference in the percentages of positive pepsin was statistically significant among 3 groups (acid, nonacid, and healthy controls):  $\chi^2 = 18.6$ ,  $P < .01$ . When the cutoff point was set as moderately positive, a sensitivity of 80% and a specificity of 85.7% were found.

### Quality of the Included Studies

The included studies ranged between 2 and 9 points regarding our quality score criteria (**Table 1**). Six of the 12 included studies were over 5 points, and the average scoring was 4.66 points. The main weakness was that authors did not clarify how much time had elapsed between performing a pepsin detection test and the gold-standard test. The second most common weakness was small sample size.

### Discussion

Over the past few years, pepsin has become a promising marker of LPR, as we can see in the growing number of studies assessing it, rendering this review highly necessary. This is the first systematic review focused exclusively on the role of pepsin as a diagnostic tool for laryngopharyngeal reflux. The available studies suggest that pepsin is a good marker for LPR. Although 2 studies are discrepant, there is a general agreement on the valuable role of pepsin as a diagnostic marker for LPR.

### Pepsin as a Diagnostic Test

In view of the evidence assessed in this study, pepsin seems to be a reliable marker for LPR. All the included studies, with the exception of 2,<sup>22,23</sup> found a relationship between LPR and pepsin.

Yadlapati et al<sup>22</sup> did not find any statistically significant differences. However, they found statistically significant differences when they performed a quantitative estimation of pepsin. Their findings could be explained because they used only RSI for the selection of patients, so there may be a possibility they had a high number of false positives. In addition, they did not define the time of day the saliva samples were taken. Other authors have highlighted the

Table 2. Description of the Included Studies.

| Author                          | City or State and Country     | Score | Sample Size   | Peptin Detection  | Sample   | Main Outcome   | Peptin in Patients   | Peptin in Controls   | Journal   | Cutoff Point  |
|---------------------------------|-------------------------------|-------|---|---|--|--|--|--|---|---|
| Fortunato et al. <sup>25</sup>  | Chicago, United States        | 7     | 90 confirmed LPR pediatric patients; 43 asymptomatic pediatric controls   | Saliva, ELISA   | Confirmed pediatric LPR patients via 24-hour combined pH-MII monitoring                        | Peptin score 3 (sensitivity = 80%, specificity = 45.7%); peptin score 4 (sensitivity = 70%, specificity = 52.8%)                             | 86% with 1 positive sample; average concentration within 30 minutes after event 38.6 ± 156 ng/mL   | 9.3% with 1 positive sample                                    | Neurogastroenterology & Motility                    | No cutoff point   |
| Yadlapati et al. <sup>22</sup>  | Illinois, United States       | 3     | 15 clinically diagnosed LPR (6 laryngeal symptoms, 9 laryngeal and esophageal symptoms); 18 asymptomatic controls | Saliva, monoclonal antibody Peptest (RD Biomed, East Yorkshire, England)  | Clinically diagnosed LPR patients (RSI and GERDq tests)  | Significant differences in quantitative estimated peptin (P = .01 and .04); no differences in percentage of positives (P = .5)               | Laryngeal symptoms patients: 40% positive; estimated peptin (7.5 ± 11.2)<br>Laryngeal and esophageal symptoms patients: 75% positive; estimated peptin (117.9 ± 147.4) | 53% positive for peptin; estimated peptin (32.4 ± 41.9)        | Clinical Gastroenterology and Hepatology            | 1.6 ng/mL   |
| Na et al. <sup>24</sup>         | Seoul, South Korea            | 9     | 50 confirmed LPR patients; 12 asymptomatic controls   | Saliva, ELISA (Cloud-Clone, Houston, Texas)   | Confirmed LPR patients via 24-hour combined pH-MII monitoring                                  | Statistically significant differences (P = .031 and .025, respectively)  | Peptin upon waking: 17.2 ng/mL<br>Peptin after lunch: 3.0 ng/mL  | Peptin upon waking: 3.7 ng/mL<br>Peptin after lunch: 0.0 ng/mL | The Laryngoscope                                    | No cutoff point   |
| lanella et al. <sup>26</sup>    | Rome, Italy                   | 3     | 20 confirmed LPR patients; 20 asymptomatic controls   | Tears, ELISA (DKG, Marburg Germany)   | Confirmed LPR pediatric patients via 24-hour combined pH-MII monitoring                        | No P value   | 20% positive for peptin  | 0% positive for peptin   | International Journal of Pediatric Otolaryngology   | 2.5 ng/mL in at least one of the eye samples<br>No cutoff value |
| Seng-Bahar et al. <sup>27</sup> | Ljubljana, Slovenia           | 6     | 28 confirmed LPR patients; 48 asymptomatic controls   | Saliva, enzyme immune test (USCN Life Science, Wuhan, China)  | Confirmed LPR patients via 24-hour combined pH-MII monitoring                                  | Statistically significant differences (P = .023)   | Total peptin µg/L: 29.8 (SD = 16)  | Total peptin µg/L: 9.3 (SD = 7.4)                              | Clinical Otolaryngology: Official Journal of ENTJUK | No cutoff value   |
| Komatsu et al. <sup>23</sup>    | Pittsburgh, United States     | 6     | 20 clinically diagnosed LPR; 10 controls (GERD patients, no LPR symptoms)   | Hypopharyngeal, gastric cardia, and distal esophagus biopsies; Western blot analysis (Abcam, Cambridge, UK)                     | Confirmed LPR patients via EGD and MII monitoring  | No statistically significant differences (P = .703, .226, and .964; for each location)   | Total peptin: 1,547.58   | Total peptin: 1,815.7  | Surgical Endoscopy                                  | No cutoff value   |
| Hayat et al. <sup>2</sup>       | London, United Kingdom        | 3     | 21 clinically diagnosed LPR; 10 asymptomatic controls   | Saliva (5 samples in 24 hours), monoclonal antibody Peptest   | Patients and controls had simultaneous MII, pH monitoring, and saliva peptin sampling          | Statistically significant differences (P = .025)   | 26.7% positive for peptin  | 17% positive for peptin  | Journal of Clinical Gastroenterology                | 5 samples; cutoff value 25 ng/mL                                |
| Birić et al. <sup>28</sup>      | Osjijek, Croatia              | 5     | 45 clinically diagnosed LPR; 30 asymptomatic controls   | Saliva, ELISA (USCN Life Science, Wuhan, China)   | Clinically diagnosed LPR patients (RSI and RFS)  | No P value was calculated  | Total peptin: 91.97 ng/mL  | Total peptin: 10.76 ng/mL                                      | International Journal of Allergology and Immunology | No cutoff value   |
| Jiang et al. <sup>29</sup>      | Canton, China                 | 5     | 15 confirmed LPR patients (7 acid, 8 nonacid); 21 asymptomatic controls   | Interarytenoid mucosal biopsies, IHQ detection of peptin, rabbit anti-human polyclonal peptin (USCN Life Science, Wuhan, China) | Confirmed LPR patients via 24-hour combined pH-MII monitoring                                  | Statistically significant differences (P = .01); no difference between acid and nonacid LPR; sensitivity of 80% and a specificity of 85.7%   | 80% positive for peptin  | 14.29% positive for peptin                                     | The Laryngoscope                                    | Moderately positive   |
| Wang et al. <sup>30</sup>       | Canton, China                 | 4     | 32 clinically diagnosed LPR; 15 asymptomatic controls   | Saliva and hypopharyngeal secretions, ELISA (USCN Life Science, Wuhan, China)   | Clinically diagnosed LPR patients (RSI, RFS, and PPI treatment tests)                          | Statistically significant differences (P = .000); no differences in peptin concentration between oral and hypopharyngeal samples (P = .136)  | 93.8% positive for peptin  | 20% positive for peptin  | Otolaryngology-Head and Neck Surgery                | No cutoff value   |
| Kim et al. <sup>31</sup>        | Suwon, Korea                  | 3     | 40 confirmed LPR patients; 12 confirmed controls  | Saliva, Western blot (Santa Cruz Biotechnology, Santa Cruz, California)   | Confirmed LPR patients via 24-hour pH monitoring; confirmed controls via 24-hour pH monitoring | No P value; no statistically significant difference in the positive rate of a peptin test between patients with and without typical symptoms | 50% positive for peptin  | 0% positive for peptin   | Digestion   | 1540 pixels, using image-pro Plus 6.0 image analysis system     |
| Johnston et al. <sup>20</sup>   | North Carolina, United States | 2     | 9 confirmed LPR patients; 12 confirmed controls   | Laryngeal biopsy (vocal fold, posterior commissure, and ventricle region), IHQ detection of peptin                              | Confirmed LPR patients via 24-hour pH monitoring; confirmed controls via 24-hour pH monitoring | Statistically significant differences (P < .001)   | 88.89% positive for peptin   | 0% positive for peptin   | The Laryngoscope                                    | No cutoff value   |

Abbreviations: EGD, esophagogastroduodenoscopy; ELISA, enzyme-linked immunosorbent assay; GERD, gastroesophageal reflux disease; GERDq, gastroesophageal reflux disease questionnaire; IHQ, immunohistochemistry; LPR, laryngopharyngeal reflux; MII, multichannel intraluminal impedance; PPI, proton pump inhibitor; RFS, Reflux Finding Score; RSI, Reflux Symptom Index.

importance of a specific time of day for saliva sample taking to detect pepsin.<sup>2,24,32</sup>

Komatsu et al<sup>23</sup> did not find a link between pepsin and LPR, but, unlike the rest of the selected studies, their control group consisted of patients with GERD, not healthy controls. Thus, it does not follow that pepsin is not related to LPR, but it calls into question if pepsin can be used to differentiate patients with GERD from patients with LPR.

In a more recent study, Yadlapati et al<sup>22</sup> addressed this very question and also found that pepsin is not useful in distinguishing between patients with LPR and GERD. In the study carried out by Kim et al,<sup>31</sup> included in this review, there were no statistically significant differences in the positive rates of pepsin tests between patients with and without typical reflux symptoms.

Therefore, according to the evidence found, pepsin does not seem to be useful in distinguishing between patients with LPR and GERD.<sup>22,23,31</sup> Future comprehensive research should be carried out.

Fortunato et al<sup>25</sup> found a low specificity rate using pepsin detection in saliva via ELISA. Other authors highlighted the low sensitivity and specificity rate for pepsin to diagnose LPR in children.<sup>33,34</sup>

### **Pepsin in Healthy Patients**

Most of the studies assessed in this review found pepsin in healthy controls, but it was at a noticeably lesser concentration than in patients (**Table 2**).

Of the 3 studies that did not find pepsin in healthy controls, 2 used confirmed controls.<sup>20,31</sup> On the other hand, the studies that found pepsin in controls used asymptomatic patients, not confirmed with the gold-standard test. On one hand, it could mean that pepsin yielded several false positives. On the other hand, it may suggest that asymptomatic LPR patients have LPR episodes.

### **Cutoff Point**

There is too much variability among selected studies in the cutoff point to consider pepsin as pathologic (**Table 2**). As some authors have highlighted before, there is no consensus<sup>30,35</sup> on this point, and it also depends on the technique used to measure pepsin. In addition, we must question the use of nonquantitative tests, such as Peptest, and whether a patient should be diagnosed with LPR with only 1 positive sample. Most of the included studies do not explain how many samples they studied, and others did not specify how many samples were studied before they considered a patient positive for pepsin.

### **Saliva, Sputum, or Biopsies as Samples**

Studies assessed in this review are divided in 2 main groups. One used saliva and sputum as samples, and the other used pharynx and larynx biopsies.

Biopsy tests seem to be more sensitive than saliva tests<sup>29,32,36</sup> but are more aggressive because they usually require sedating the patient. Most authors took biopsies from unconscious patients. However, in one of the assessed studies in this review, biopsies were taken from conscious patients, with the area numbed by local anesthesia.<sup>29</sup> This

technique could be widely used in otolaryngology examination rooms. On the other hand, saliva/sputum tests are less aggressive. Therefore, they are more easily accepted by patients and also accessible to all practitioners.

Another problem among investigations that studied biopsies is that there is no consensus about which area to take the biopsy from. The selected studies include the postcricoid area, interaritenoid mucosa, vocal fold, ventricle, posterior commissure, and hypopharynx.<sup>20,29,37,38</sup>

The other main group of studies studied pepsin in saliva/sputum. Some studies in this group used the same diagnostic tool, the Peptest commercial kit.<sup>2,22</sup> The other portion of these studies used IHQ techniques such as ELISA or Western blot.<sup>24-28,30,31</sup> These techniques are much more expensive and unavailable in most facilities; however, they have the advantage of being quantitative tools.

Another important difference of this group is the number of samples and the time of day they were taken. In a work included in this review, Fortunato et al<sup>25</sup> found a wide range of pepsin values throughout the day in patients with LPR. The most common times are morning, symptomatic episodes, and night. Taking the sample upon the patient waking<sup>24</sup> or immediately after the reflux episode<sup>25</sup> seems to be the most useful. Some authors used more samples, and they found that the more samples they had, the more patients they diagnosed.<sup>2</sup>

Although the results of this review offer strong evidence for the use of pepsin as a marker of LPR, the role it plays in the diagnostic process remains unclear, as well as the best way to measure it. Therefore, research should be carried out to compare the Peptest with biopsies to clarify which method is better in the diagnostic process.

### **Conclusions**

To sum up, systematic review of the current literature about pepsin suggests that it might be a reliable marker in patients with LPR, although questions remain about optimal timing, location, nature, and threshold values for pepsin testing.

### **Authors' Note**

This work is part of the research completed by Christian Calvo-Henríquez, MD, to obtain a PhD degree.

### **Author Contributions**

**Christian Calvo-Henríquez**, data analysis, drafting, final approval, accountability for all aspects of the work; **Alberto Ruano-Ravina**, data analysis, drafting, final approval, accountability for all aspects of the work; **Pedro Vaamonde**, data analysis, drafting, final approval, accountability for all aspects of the work; **Gabriel Martínez-Capoccioni**, data analysis, drafting, final approval, accountability for all aspects of the work; **Carlos Martín-Martín**, data analysis, drafting, final approval, accountability for all aspects of the work.

### **Disclosures**

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## References

1. Koufman JA. The otolaryngologic manifestations of gastroesophageal reflux disease (GERD): a clinical investigation of 225 patients using ambulatory 24-hour pH monitoring and an experimental investigation of the role of acid and pepsin in the development of laryngeal injury. *Laryngoscope*. 1991;101(4, pt 2)(suppl 53):1-78.
2. Hayat JO, Yazaki E, Moore AT, et al. Objective detection of esophagopharyngeal reflux in patients with hoarseness and endoscopic signs of laryngeal inflammation. *J Clin Gastroenterol*. 2014;48:318-327.
3. Vaezi MF, Schroeder PL, Richter JE. Reproducibility of proximal probe pH parameters in 24-hour ambulatory esophageal pH monitoring. *Am J Gastroenterol*. 1997;92:825-829.
4. Reichel O, Issing WJ. Impact of different pH thresholds for 24-hour dual probe pH monitoring in patients with suspected laryngopharyngeal reflux. *J Laryngol Otol*. 2008;122:485-489.
5. Belafsky PC, Postma GN, Koufman JA. Validity and reliability of the reflux symptom index (RSI). *J Voice*. 2002;16:274-277.
6. Belafsky PC, Postma GN, Koufman JA. The validity and reliability of the reflux finding score (RFS). *Laryngoscope*. 2001;111:1313-1317.
7. Karkos PD, Wilson JA. Empiric treatment of laryngopharyngeal reflux with proton pump inhibitors: a systematic review. *Laryngoscope*. 2006;116:144-148.
8. Park KH, Choi SM, Kwon SUK, Yoon SW, Kim SUK. Diagnosis of laryngopharyngeal reflux among globus patients. *Otolaryngol Head Neck Surg*. 2006;134:81-85.
9. Branski RC, Bhattacharyya N, Shapiro J. The reliability of the assessment of endoscopic laryngeal findings associated with laryngopharyngeal reflux disease. *Laryngoscope*. 2002;112:1019-1024.
10. Vaezi MF. Gastroesophageal reflux disease and the larynx. *J Clin Gastroenterol*. 2003;36:198-203.
11. Cohen SM, Kim J, Roy N, Courey M. Prescribing patterns of primary care physicians and otolaryngologists in the management of laryngeal disorders. *Otolaryngol Head Neck Surg*. 2013;149:118-125.
12. Francis DO, Rymer JA, Slaughter JC, et al. High economic burden of caring for patients with suspected extraesophageal reflux. *Am J Gastroenterol*. 2013;108:905-911.
13. Vaezi MF, Hicks DM, Abelson TI, Richter JE. Laryngeal signs and symptoms and gastroesophageal reflux disease (GERD): a critical assessment of cause and effect association. *Clin Gastroenterol Hepatol*. 2003;1:333-344.
14. Koufman JA, Aviv JE, Casiano RR, Shaw GY. Laryngopharyngeal reflux: position statement of the committee on speech, voice, and swallowing disorders of the American Academy of Otolaryngology—Head and Neck Surgery. *Otolaryngol Head Neck Surg*. 2002;127:32-35.
15. Johnston N, Dettmar PW, Bishwokarma B, Lively MO, Koufman JA. Activity/stability of human pepsin: implications for reflux attributed laryngeal disease. *Laryngoscope*. 2007;117:1036-1039.
16. Johnston N, Wells CW, Blumin JH, Toohill RJ, Merati AL. Receptor-mediated uptake of pepsin by laryngeal epithelial cells. *Ann Otol Rhinol Laryngol*. 2007;116:934-938.
17. Johnston N, Wells CW, Samuels TL, Blumin JH. Rationale for targeting pepsin in the treatment of reflux disease. *Ann Otol Rhinol Laryngol*. 2010;119:547-558.
18. Johnston N, Dettmar PW, Lively MO, et al. Effect of pepsin on laryngeal stress protein (Sep70, Sep53, and Hsp70) response: role in laryngopharyngeal reflux disease. *Ann Otol Rhinol Laryngol*. 2006;115:47-58.
19. Samuels TL, Johnston N. Pepsin as a causal agent of inflammation during nonacidic reflux. *Otolaryngol Head Neck Surg*. 2009;141:559-563.
20. Johnston N, Knight J, Dettmar PW, Lively MO, Koufman J. Pepsin and carbonic anhydrase isoenzyme III as diagnostic markers for laryngopharyngeal reflux disease. *Laryngoscope*. 2004;114:2129-2134.
21. Vaezi MF, Slaughter JC, Smith BS, et al. Dilated intercellular space in chronic laryngitis and gastro-oesophageal reflux disease: at baseline and post-lansoprazole therapy. *Aliment Pharmacol Ther*. 2010;32:916-924.
22. Yadlapati R, Adkins C, Jaiyeola DM, et al. Abilities of oropharyngeal pH tests and salivary pepsin analysis to discriminate between asymptomatic volunteers and subjects with symptoms of laryngeal irritation. *Clin Gastroenterol Hepatol*. 2016;14:535-542.e2.
23. Komatsu Y, Kelly LA, Zaidi AH, et al. Hypopharyngeal pepsin and Sep70 as diagnostic markers of laryngopharyngeal reflux: preliminary study. *Surg Endosc*. 2015;29:1080-1087.
24. Na SY, Kwon OE, Lee YC, Eun YG. Optimal timing of saliva collection to detect pepsin in patients with laryngopharyngeal reflux. *Laryngoscope*. 2016;126:2770-2773.
25. Fortunato JE, D'Agostino RB, Lively MO. Pepsin in saliva as a biomarker for oropharyngeal reflux compared with 24-hour esophageal impedance/pH monitoring in pediatric patients [published online September 7, 2016]. *Neurogastroenterol Motil*.
26. Iannella G, Di Nardo G, Plateroti R, et al. Investigation of pepsin in tears of children with laryngopharyngeal reflux disease. *Int J Pediatr Otorhinolaryngol*. 2015;79:2312-2315.
27. Sereg-Bahar M, Jerin A, Jansa R, Stabuc B, Hocevar-Boltezar I. Pepsin and bile acids in saliva in patients with laryngopharyngeal reflux—a prospective comparative study. *Clin Otolaryngol*. 2015;40:234-239.
28. Birtić D, Vceva A, Kotromanović Z, Zubčić Z, Mihalj H, Jovanović S. Significance of the pepsin from the saliva in the diagnosis and treatment of laryngopharyngeal reflux disease. *Coll Antropol*. 2012;36(suppl 2):83-86.
29. Jiang A, Liang M, Su Z, et al. Immunohistochemical detection of pepsin in laryngeal mucosa for diagnosing laryngopharyngeal reflux. *Laryngoscope*. 2011;121:1426-1430.
30. Wang L, Liu X, Liu Y, et al. Correlation of pepsin-measured laryngopharyngeal reflux disease with symptoms and signs. *Otolaryngol Head Neck Surg*. 2010;143:765-771.
31. Kim TH, Lee KJ, Yeo M, Kim DK, Cho SW. Pepsin detection in the sputum/saliva for the diagnosis of gastroesophageal reflux disease in patients with clinically suspected atypical gastroesophageal reflux disease symptoms. *Digestion*. 2008;77:201-206.
32. Saritas Yuksel E, Hong S-KS, Strugala V, et al. Rapid salivary pepsin test: blinded assessment of test performance in gastroesophageal reflux disease. *Laryngoscope*. 2012;122:1312-1316.

33. Safe M, Cho J, Krishnan U. Combined multichannel intraluminal impedance and pH measurement in detecting gastroesophageal reflux disease in children. *J Pediatr Gastroenterol Nutr.* 2016;63:e98-e106.
34. Dy F, Amirault J, Mitchell PD, Rosen R. Salivary pepsin lacks sensitivity as a diagnostic tool to evaluate extraesophageal reflux disease. *J Pediatr.* 2016;177:53-58.
35. Hayat JO, Gabieta-Somnez S, Yazaki E, et al. Pepsin in saliva for the diagnosis of gastro-oesophageal reflux disease. *Gut.* 2015;64:373-380.
36. Knight J, Lively MO, Johnston N, Dettmar PW, Koufman JA. Sensitive pepsin immunoassay for detection of laryngopharyngeal reflux. *Laryngoscope.* 2005;115:1473-1478.
37. Wassenaar E, Johnston N, Merati A, et al. Pepsin detection in patients with laryngopharyngeal reflux before and after fundoplication. *Surg Endosc.* 2011;25:3870-3876.
38. Gill GA, Johnston N, Buda A, et al. Laryngeal epithelial defenses against laryngopharyngeal reflux: investigations of E-cadherin, carbonic anhydrase isoenzyme III, and pepsin. *Ann Otol Rhinol Laryngol.* 2005;114:913-921.